

STEREOCHEMISTRY OF TRITIUM AT C-1 AND C-7
IN CHOLESTEROL DERIVED FROM (3R,2R)-
2T-MEVALONIC ACID

E. Caspi, J. B. Greig, P. J. Ramm and K. R. Varma
Worcester Foundation for Experimental Biology
Shrewsbury, Massachusetts

(Received in USA 25 April 1968; received in UK for publication 4 June 1968)

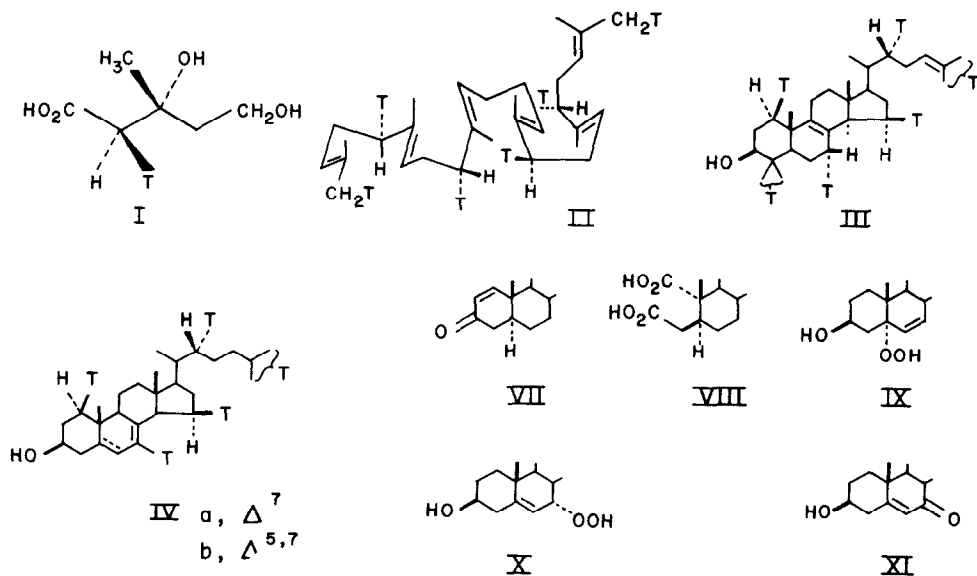
Consideration of the incorporation of mevalonic acid stereospecifically labeled with isotopic hydrogen at C-2, into squalene⁽¹⁾ and the cyclisation of the latter into sterols allows the prediction of the stereochemistry of the tritium atoms in the biosynthesised products. Hence, (3R,2R)-2T-mevalonic acid (I) leads to 1, 5R, 9R, 16R, 20R, 24 T_G-squalene (II). Cyclisation of this squalene will give T_G-lanosterol (III) labeled in the 1β, 7α, 15β, 22R, 30 or 31 and 26 or 27 positions. The transformation from lanosterol to cholesterol involves the intermediacy⁽²⁾ first of a C-7 olefin (IVa) and then a 5, 7 diene (IVb). Consequently, one of the C-7 protons which originate from C-2 of mevalonic acid will be lost during the transformation⁽³⁾. In this communication we concern ourselves with the fate and the stereochemistry of the tritium atoms at C-1 and C-7 in cholesterol derived from (3R,2R)-2T-mevalonic acid.

Cholesterol was biosynthesised⁽⁴⁾ by incubation of (3R,2R)-2T-2¹⁴C-mevalonic acid with a rat liver preparation⁽⁵⁾. The cholesterol was purified by thin layer chromatography in two systems (a) ethyl acetate: hexane 3:7 and (b) methanol:benzene 1:19. A portion of the material (1.72 x 10⁶ d.p.m. ¹⁴C)⁽⁶⁾ was diluted with non-radioactive cholesterol (1.5 g), and crystallized to constant specific activity giving a T/¹⁴C ratio of 10.1.

TABLE
 $T/^{14}C$ Ratios* And Number of Tritium Atoms

	$T/^{14}C$	No. T. Atoms
Cholesterol	10.1	5.00
Cholest-4-en-3-one (V)	9.7	4.80
5 α -Cholestan-3-one (VI)	9.7	4.80
5 α -Cholest-1-en-3-one (VII)	10.0	4.95
1,2-seco-5 α -cholestan-1,3-dioic acid (VIII)	8.2	4.06
5 α -Hydroperoxycholest-6-en-3 β -ol (IX)	9.6	4.85
7 α -Hydroperoxycholest-5-en-3 β -ol (X)	9.7	4.80
Cholest-5-en-3 β -ol-7-one	8.1	4.01

*The $T/^{14}C$ ratios were determined on recrystallized materials and were constant. The results represent the averages of nine determinations. The number of T atoms is calculated on the basis of the $T/^{14}C$ (10.1) ratio for cholesterol.



For the determination of the presence and stereochemistry of tritium at C-1, the cholesterol was oxidised by the Oppenauer method⁽⁷⁾ to cholest-4-en-3-one (V) ($T/^{14}C$ ratio 9.7). (Table). Reduction of V with lithium in liquid ammonia⁽⁸⁾ gave 5 α -cholestan-3-one (VI) ($T/^{14}C$ ratio 9.7). Dehydrogenation of VI with dichlorodicyanobenzoquinone (DDQ)⁽⁹⁾ yielded 5 α -cholest-1-en-3-one (VII) m.p. 109-110^o ($T/^{14}C$ ratio 10.0). Finally, oxidative cleavage of VII gave 1,2-seco-5 α -cholestan-1,3-dioic acid (VIII) m.p. 216.5-216.5^o (m/e:420, M⁺): ($T/^{14}C$ ratio 8.2). The 1/5 decrease in $T/^{14}C$ ratio reflects the loss of one out of five tritium atoms, during the oxidation, this tritium being located at C-1. Since Ringold and Turner⁽⁹⁾ demonstrated that DDQ dehydrogenation of steroidal 5 α -3-ketones proceeds with loss of the 1 α and 2 β axial hydrogens, then the tritium atom at C-1 must have the expected 1 β -stereo-chemistry.

The demonstrated presence of five tritium atoms in the cholesterol indicates that isotopic hydrogen is still present at C-7. It could therefore be inferred that the formation of the Δ^7 intermediate (IVa) involves the elimination⁽¹⁰⁾ of the 7 β rather than the 7 α hydrogen. Confirmation of the presence of a tritium atom at C-7 and of its stereochemistry was sought and is now described.

Cholesterol was photolysed in the presence of oxygen and hematoporphyrin⁽¹¹⁾ giving 5 α -hydroperoxycholest-6-en-3 β -ol (IX) m.p. 147-149^o ($T/^{14}C$ ratio 9.8). Isomerisation^(12,13) of IX gave 7 α -hydroperoxycholest-5-en-3 β -ol (X) m.p. 154-156^o ($T/^{14}C$ ratio 9.7). Simultaneous isomerisation and dehydration of IX with cupric chloride⁽¹²⁾ in pyridine gave cholest-5-en-3 β -ol-7-one (XI) ($T/^{14}C$ ratio 8.1). The 1/5 decrease in $T/^{14}C$ ratio again indicates the loss of one out of five isotopic hydrogens and proves unequivocally the presence of a tritium atom at C-7. Since the photooxidation of cholesterol is a stereospecific process involving the loss of the 7 α hydrogen atom⁽¹¹⁾, the tritium must have the 7 β configuration. It follows that the saturation of Δ^7 double bond of the 5,7-diene (IVb) involves the addition, at C-7, of a hydrogen which assumes the α -stereo-chemistry⁽¹⁴⁾. The overall result of the events occurring at C-7 during the conversion of lanosterol to cholesterol is therefore the inversion from the 7 α to the 7 β configuration of the proton originating from the pro-2R hydrogen of mevalonic acid.

Acknowledgement This work was supported by grant P-500-G from the American Cancer Society and grant K3-16614 from the National Cancer Institute.

REFERENCES

1. G. Popjak and J. W. Cornforth, *Biochem. J.* 101, 553 (1966).
2. For pertinent references see I. D. Frantz and G. J. Schroepfer, *Ann. Rev. Biochem.* 36, 691 (1967)
3. L. Canonica, A. Fiecchi, M. Galli Kienle, A. Scala, G. Galli, E. Grossi Paoletti and R. Paoletti, *Steroids* 11, 287 (1968)
4. Results for the biosynthetic intermediates squalene and lanosterol will be published at a later date.
5. J. W. Cornforth, R. H. Cornforth, A. Pelter, M. G. Horning and G. Popjak, *Tetradedon* 5, 311 (1959).
6. Samples were counted on a Nuclear-Chicago, automatic liquid scintillation counter, Model Mark I. The samples were dissolved in 15 ml of a scintillator solution of toluene containing 4 g of 2,5-diphenyloxazole and 100 mg of p-bis-2-(5-phenyloxazolyl)-benzene, per 1000 ml. The observed variations in the T/¹⁴C ratios of all compounds described fall within experimental error of the method employed.
7. The physical constants and spectroscopic results (i.r., n.m.r. and mass spectra) were consistent with the assigned structures in all cases.
8. F. L. Weisenborn and H. E. Applegate, *J. Amer. Chem. Soc.*, 81, 1960 (1959).
9. A. B. Turner and H. J. Ringold, *J. Chem. Soc. (c)*, 1720 (1967).
10. M. Akhtar and A. D. Rahimtula, *Chem. Comm.*, 259 (1968).
11. A. Nickon and J. F. Bagli, *J. Amer. Chem. Soc.*, 83, 1498 (1961).
12. G. O. Schenk, O. A. Neumuller and W. Eisfeld, *Ann.*, 616 202 (1958).
13. B. Lythgoe and S. Tripett, *J. Chem. Soc.*, 471 (1959).
14. D. C. Wilton, K. A. Munday, S. J. M. Skinner and M. Akhtar, *Biochem. J.*, 106, 603 (1968).